

**Research Article** 

# Invasion of Asian tiger shrimp, *Penaeus monodon* Fabricius, 1798, in the western north Atlantic and Gulf of Mexico

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#### Abstract

After going unreported in the northwestern Atlantic Ocean for 18 years (1988 to 2006), the Asian tiger shrimp, *Penaeus monodon*, has recently reappeared in the South Atlantic Bight and, for the first time ever, in the Gulf of Mexico. Potential vectors and sources of this recent invader include: 1) discharged ballast water from its native range in Asia or other areas where it has become established; 2) transport of larvae from established non-native populations in the Caribbean or South America via ocean currents; or 3) escape and subsequent migration from active aquaculture facilities in the western Atlantic. This paper documents recent collections of *P. monodon* from the South Atlantic Bight and the Gulf of Mexico, reporting demographic and preliminary phylogenetic information for specimens collected between North Carolina and Texas from 2006 through 2012. The increased number of reports in 2011 and 2012, ranging from 102 mm to 298 mm total length, indicates that an adult population is present in densities sufficient for breeding, which is indicative of incipient establishment. Based on these reports of *P. monodon*, its successful invasion elsewhere, and its life history, we believe that this species will become common in the South Atlantic Bight and Gulf of Mexico in less than 10 years. *Penaeus monodon* is an aggressive predator in its native range and, if established, may prey on native shrimps, crabs, and bivalves. The impacts of an established *P. monodon* population are potentially widespread (e.g., alterations in local commercial fisheries, direct and indirect pressures on native shrimp, rab and bivalve populations, and subsequent impacts on the populations of other predators of those organisms) and should be considered by resource managers. The impacts of *P. monodon* on native fauna and the source(s) or vector(s) of the invasion, however, remain unknown at this time.

Key words: Asian tiger shrimp, Penaeus monodon, western Atlantic, Gulf of Mexico, invasion, population status, phylogenetics

### Introduction

The Asian tiger shrimp, *Penaeus monodon*, is a widespread penaeid shrimp species that is native to the Indo-West Pacific (Figure 1), with a range comprising southern Japan, Korea, China, Taiwan, the Philippines, Vietnam, Cambodia, Malaysia, Singapore, Indonesia, Papua New Guinea, Australia, Thailand, Myanmar, Bangladesh, Sri Lanka, India, Pakistan, Tanzania, Madagascar, and South Africa (Motoh 1981; FAO 2012), and the Red Sea off Yemen (US National Museum of Natural History Cat. No. 171584). *Penaeus monodon* has been widely farmed outside of its native range,

including West Africa and various locations in the western Atlantic. This species is now established in many areas due to escapes from aquaculture including West Africa (Sahel and West Africa Club 2006; Ayinla et al. 2009; Anyanwu et al. 2011; Global Biodiversity Information Facility 2013), the Caribbean (Gómez-Lemos and Campos 2008), and along the northern and northeastern coasts of South America from Venezuela to eastern Brazil (e.g., Coelho et al. 2001; Silva et al. 2002; Aguado and Sayegh 2007; Cintra et al. 2011). *Penaeus monodon* has also been introduced to Hawaii, Tahiti, and England (Rodríguez and Suárez 2001), but is not established in these locations. There

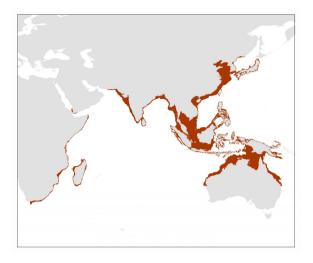


Figure 1. Native distribution of *Penaeus monodon* (adapted from FAO 2012).

are recent unconfirmed reports of shrimpers catching *P. monodon* in Haiti and a single confirmed report from the Dominican Republic in 2006 (A. Stokes, South Carolina Department of Natural Resources, Waddell Mariculture Center, Bluffton, SC, pers. comm.). The first occurrence in Puerto Rico was reported in June 2012 (USGS 2013), and an August 2012 report from Jamaica was also verified (D. Buddo, University of West Indies, Discovery Bay Marine Laboratory, Kingston, Jamaica, pers. comm.). There was an attempt to culture P. monodon in Florida in 2004 (P. Zajicek, Florida Department of Agriculture and Consumer Services, Tallahassee, FL, pers. comm.) but it was unsuccessful and the facility subsequently closed.

In the summer of 1988, an unknown number of P. monodon were accidentally released from a culture pond in South Carolina (A. Stokes, pers. comm.; D. Whitaker, South Carolina Department of Natural Resources, Marine Resources Division, Charleston, SC, pers. comm.). Nearly 300 of these shrimp were subsequently collected in trawl nets off the coasts of South Carolina, Georgia, and northeastern Florida in the following two months (D. Knott, pers. comm.) with no additional reports during the following 18 years. In September 2006, a single adult male was captured in the Mississippi Sound near Dauphin Island, Alabama (L. Hartman, pers. comm.). One month later, five specimens were collected in Pamlico Sound, North Carolina (T. Moore, North Carolina Department of Environment and Natural Resources, Mooresville,

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NC, pers. comm.). In late summer 2007, a single specimen was caught in Vermilion Bay, Louisiana (H. Blanchet, Louisiana Department of Wildlife and Fisheries, Baton, Rouge, LA, pers. comm.), and several more were reported from North Carolina, South Carolina, and Florida. The first verified collections of P. monodon in Georgia, Mississippi, and Texas occurred in 2008, 2009, and 2011, respectively. To date, Penaeus monodon has been documented in the U.S. from North Carolina to Texas (Figure 2), with many locations reporting multiple collections per year. In all cases, only a few specimens of P. monodon were caught. In October 2013, however, two commercial shrimpers reported catches of 11 and 18 kg of P. monodon in single trips off Flagler Beach, Florida. Clearly at least one breeding population of P. monodon now exists in eastern waters of the USA.

Following the 2006 reappearance of P. monodon in the waters of the South Atlantic Bight and the Gulf of Mexico, researchers from state and federal agencies and independent scientists came together, under the auspices of the Gulf and South Atlantic Regional Panel (GSARP) of the Aquatic Nuisance Species Task Force to: 1) review information on the biology of *P. monodon*, in order to make informed predictions of economic and ecological impacts; and 2) coordinate information on reports of recreational and commercial catches of *P. monodon* within the waters covered by the GSARP, namely the U.S. coastline from North Carolina to Texas. In this paper, we present demographic data (distribution, size, weight, and sex ratio) of the animals collected between 2006 and 2012, highlight the pertinent aspects of the biology of P. monodon related to this invasion, and discuss preliminary phylogenetic information on the animals collected between 2006 and 2012.

### Methods

### Species biology and life history

Estuaries serve as nursery grounds for *P. monodon* (Mohamed 1967; Chaudhari and Jalihal 1993), with larvae, juveniles, and young sub-adults occupying shallow coastal estuaries, lagoons, and mangrove areas. Sub-adults subsequently move offshore, where they are typically found in depths up to 70 m, although they are known to occur in water as deep as 162 m (Motoh 1981). *P. monodon* matures and breeds predominantly on sand or muddy-sand bottom in these nearshore marine habitats.

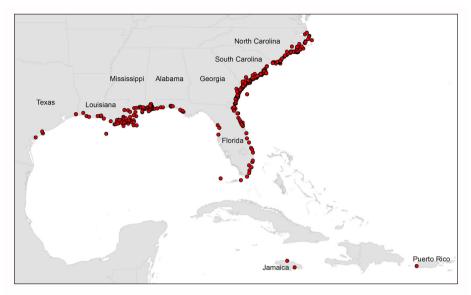


Figure 2. Map showing reported Penaeus monodon collections in the U.S. from 2006–2012 (USGS 2013).

Females grow larger than males (Primavera et al. 1998) and can reach 330 mm total length (hereafter TL, measured from tip of rostrum to tip of telson) (FAO 2013), with the largest specimen recorded being 337 mm TL (Crosnier 1965 in Mohamed 1967). The maximum size of males reported by Motoh (1985) is 71 mm carapace length (hereafter CL), which is equivalent to ~240 mm TL (Primavera et al. 1998). In wild populations, males become sexually mature at an age of  $\sim 5$  months and a body weight of  $\sim 35$  g (~32 mm CL  $\approx$ 150 mm TL). Females are capable of breeding at ~6 months and a body weight of ~70 g (~45–50 mm CL  $\approx$ 175 mm TL) (Primavera et al. 1998; FAO 2013). In one particular pond culture setting, P. monodon size and age were related as follows: 4 months (120 mm TL, 14 g), 6 months (142 mm TL, 22 g), 10 months (211 mm TL, 63 g), 12 months (229 mm TL, 95 g) (Delmendo and Rabanal 1956 in Mohamed 1967). The difference between body size and age for wild versus cultured shrimp implies that, at least in this culture situation (Delmendo and Rabanal 1956), wild animals grow faster than cultured ones.

Male and female *P. monodon* live 1.5 and 2 years, respectively, based on pond rearing experiments in the Philippines (Motoh 1981). A shorter life span of 12–14 months was determined for wild penaeids (including *P. monodon*), in western India (Srivatsa 1953 in Mohamed 1967).

The timing of spawning activity has been studied in Singapore and the Philippines. Hall

(1962) suggested that in the waters off Singapore, P. monodon breeds between February and April. In the Philippines, there is limited spawning year round, but there are peaks in March, and October or November depending on the location (Motoh 1981). Female P. monodon are highly fecund, producing between 200,000 and 1 million eggs in a complete spawn with an average of ~500,000 (Primavera 1982). Following spawning, both pondreared and wild-caught females may re-mature and repeat spawning, with large individuals doing so more frequently and producing more eggs than small ones (Menasveta et al. 1994). Pond-reared females of 50-63 mm CL, with one evestalk removed, spawned viable eggs repeatedly within inter-molt periods of 20-30 days, indicating that one impregnation was sufficient to fertilize several batches of eggs spawned within one inter-molt period (Beard and Wickins 1980).

*Penaeus monodon* tolerate salinities from 0 to as high as 38 psu (Motoh 1981; Chaudhari and Jalihal 1993). Lethal thermal extremes are not definitively known, although mortality has been reported at water temperatures below 13 °C and above 33 °C (Jintoni 2003) and below 10 °C or above 39 °C for postlarvae and juveniles (Motoh 1981). Survival and growth, however, are severely limited below 20 °C (Lumare et al. 1993).

A primary concern regarding the impacts of the introduction of *P. monodon* outside of its native range is the potential to compete with, or





**Figure 3.** *Top*: Lateral view of a mature female *Penaeus monodon* (279 mm TL) caught in 2008 off Charleston Harbor, SC (photo courtesy of the SCDNR Southeastern Regional Taxonomic Center, SERTC). *Bottom*: An immature "red-striped" specimen (152 mm TL) caught by cast net in April 2012 in the Intracoastal Waterway near Vero Beach, FL (photo courtesy of Tom Stokes).

prey directly upon, native species. *Penaeus* monodon preys upon young penaeid prawns (Thomas 1972); a wide variety of macroinvertebrates (e.g., gastropods, bivalves, crustaceans, and polychaetes), fish, as well some plant material, and small amounts of echinoderms, hydroids, debris, silt, and sand (Marte 1980; Luna-Marte 1982; Dall 1992; Smith et al. 1992). At least one study suggests that they do not feed upon carrion (Hill and Wassenberg 1987).

### Identification

Mature *Penaeus monodon* can be distinguished from native penaeid shrimp of the southeastern U.S. by their large size (up to 330 mm TL and 330 g in total wet weight, hereafter WT), their overall rusty brown to black color, and/or by the distinctive dark and light alternating banding across the back of the thorax and abdomen (Figure 3, top panel) (Mohamed 1967). Although the lighter bands are sometimes indistinct on juvenile shrimp (e.g.,  $\sim 100-150$  mm TL), yellow patches near the bases of the percopods and pleopods and yellow banding on the antennules and antennae can be useful in distinguishing *P. monodon* from penaeids native to the U.S. (Knott, pers. obs.). Aside from their characteristic coloration, juveniles and sub-adults of *P. monodon* can be differentiated from native species by the posterior extent of the adrostral sulci and the morphology of the external genitalia (Motoh and Buri 1980; Pérez Farfante and Kensley 1997).

There is also a color variant of P. monodon that has a conspicuous, wide, reddish-orange stripe along the dorsum that extends from the rostrum to the telson (Figure 3, bottom panel). This "red-striped" morph expresses an allele that is present in < 1% of wild populations in Australia; however, lineages expressing the trait in ~60% of individuals in some families have been derived through selective breeding of animals taken from Australian populations (J. Aquaculture Wyban, High Health Inc.. Kurtistown, HI, pers. comm.). A low proportion (6.6%) of the *P. monodon* reported in the U.S. in 2012 showed this trait; however, a much greater proportion (52.9%) of individuals  $\leq 152 \text{ mm TL}$ exhibited this trait, suggesting that it may be an age- or size-related characteristic.

## *Gathering information on reports of* Penaeus monodon *in the U.S.*

Regional coordinators from North Carolina to Texas distributed information and coordinated P. monodon reports from recreational and commercial fishermen. Most state agencies between North Carolina and Texas conducted publicity campaigns in the form of press releases and the distribution of flyers and posters that informed the public that collection information and specimens were being sought to investigate the P. monodon invasion. Outreach information was dispersed freely with some coordination on the content and presentation. In cases where the specimens were given to biologists, TL, WT, and gender were recorded for each specimen collected, along with precise information on collection location (e.g., latitude and longitude, if available). Females were determined by the presence of a thelycum, an external reproductive opening; males by the presence of a petasma (Mohamed 1967; Motoh 1981). None of the females were examined for eggs. Persons collecting P. monodon were also encouraged to submit photographs of specimens as part of their reports

Year	NC	SC	GA	FL	AL	MS	LA	ТХ	Total
2005	0	0	0	0	0	0	0	0	0
2006	5	0	0	0	1	0	0	0	6
2007	1	1	0	1	0	0	1	0	4
2008	8	6	4	2	1	0	0	0	21
2009	14	15	3	1	5	3	4	0	45
2010	2	20	1	2	0	0	7	0	32
2011	329	144	3	25	28	16	128	5	678
2012	21	64	43	41	3	13	9	1	195
Total	380	250	54	72	38	32	149	6	981

Table 1. Number of individuals of Penaeus monodon collected by state and year from 2005–2012.

in order to confirm species identification. In some cases, objects in these photographs (e.g., coins, rulers, newspaper headlines, etc.) were used to determine specimen size when measurements were not provided directly or their accuracy was questionable.

### Statistical analyses

In order to examine the effects of sex and length on weight and the interaction between sex and length, an ANCOVA was performed on  $\log_{10}$ - $\log_{10}$  transformed TL and WT using both the entire data set and also separately on the subset of data for *P. monodon* from 150–250 mm TL. Any eggs that may have been present in females were not removed. All statistical analyses were performed using Minitab version 16.2.2.

### Results

As of December 31, 2012, the distribution of *P. monodon* in coastal and estuarine waters of the southeastern U.S. extended from Albemarle Sound, North Carolina on the Atlantic U.S. coast to Aransas Bay, Texas in the western Gulf of Mexico, as well as a few locations in the eastern Caribbean (Figure 2). A detailed account of reports that define this distribution can be found in the validated collections of specimens in the USGS Nonindigenous Aquatic Species database (USGS 2013).

Following initial reports of small numbers of *P. monodon* in 2006 and 2007, the number of reported specimens increased slightly between 2008 and 2010 (Table 1, Figure 4) and was followed by a 20-fold increase in reports in 2011. Some of this increase may be attributable to increased levels of public outreach and widespread media coverage; however, numerous

shrimp fishermen described catching P. monodon with increased frequency and in greater numbers, suggesting that their actual abundance increased substantially in 2011. The primary source of P. monodon reports was direct communication with commercial shrimpers who were actively fishing in the Gulf of Mexico and the South Atlantic Bight. In 2012, a number of commercial fishermen informed dock personnel that *P. monodon* had become more abundant than in the previous year, but in some cases specimens were eaten or sold without being reported. Fisheries-independent and catch-per-unit-effort data are not available to examine how well the reported occurrences reflect changes in the actual abundance of this invasive species since its arrival in coastal U.S. waters.

By region, the number of *P. monodon* reported from the Gulf of Mexico is lower than that in the South Atlantic Bight. From 2006-2012, 737 P. monodon were captured along the Atlantic coast, from Albemarle Sound, North Carolina through the Florida Keys. The greatest number (~85%) was caught by commercial shrimp fishermen trawling in North and South Carolina (Table 1). During that same period in the Gulf of Mexico, 244 specimens were captured, with all but one individual (from the Dry Tortugas National Park off southwest Florida Keys) being captured between Clearwater, Florida and Aransas Bay, Texas (Figure 2). More than 60% of these P. monodon from the Gulf of Mexico were captured in the waters of Louisiana (Table 1).

Among those specimens for which a definitive month of capture was recorded, by far the greatest number (>90%) of *P. monodon* was captured during the months of August through November (Figure 5). This peak coincided with the latter half of the most intense commercial shrimp fishing effort (NOAA 2013). The 3–6 month lag

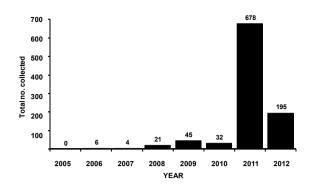


Figure 4. Number of *Penaeus monodon* collected by year from 2005–2012.

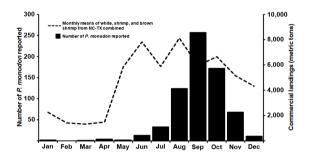


Figure 5. Number of *Penaeus monodon* collected by month from 2006–2012 and monthly average (in metric tons) of south Atlantic and Gulf coast state's total commercial landings of native white, pink and brown shrimp from 2006–2011 (latest available data from query of NOAA's NMFS Monthly Commercial Landings Statistics database).

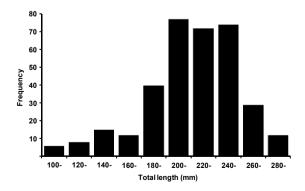


Figure 6. Length-frequency distribution of *Penaeus monodon* collected in U.S. waters from 2006-2012 (total n = 345).

time between the seasonal increase in shrimp landings and the peak period of *P. monodon* collection suggests that the peak is not merely a reflection of commercial shrimp fishing effort (Figure 5) and may be a result of reduced *P. monodon* detection at smaller sizes.

Individuals ranged in size from 102 mm to 298 mm TL, with the majority being adults between 200 mm and 259 mm TL (Figure 6). The occurrence of smaller individuals (juveniles) strongly suggests that P. monodon may be reproducing within or close to waters of the South Atlantic Bight and/or the Gulf of Mexico. Among the shrimp caught between April and August, the number of juveniles (< 160 mm TL) increased more than two-fold between 2011 and 2012 (Figure 7). Despite the fact that overall fewer P. monodon were collected in 2012 (Figure 4), the greatest abundance of small P. monodon was observed during that year (Figure 7d). Among P. monodon specimens for which both the date of collection and size were available, those captured in the late spring and early (April-August) were smaller, summer on average, than those caught later in the year (September-January) during the period of peak shrimp trawling (195.6 mm TL, n = 72 vs. 226.7 mm TL, n = 273, respectively; t = -5.45, P <0.001).

Although the vast majority of P. monodon (89%) were captured by commercial fishermen, a seasonal difference was observed in the type of gear used by reporters. Approximately 20% of the reports from April through August (n = 142)were made by individuals using gear other than commercial trawl nets, principally cast nets and crab pots. For the following months (September through January), the percentage of reports where non-commercial gear was used decreased to 6% (n = 281). This decline likely reflects the appreciable elevation in commercial fishing pressure during the fall and early winter months. It is possible, however, that some of the difference could also be attributed to P. monodon occurring at greater depths during this period, making them less likely to be captured by non-trawling methods.

The greatest increase in *P. monodon* size (TL) occurred between May and August, followed by relatively little change through the fall and early winter (Figure 8). As a proxy for growth, this amounts to an average increase of 9–10 mm TL per month for combined males and females collected between May and January of the following year.

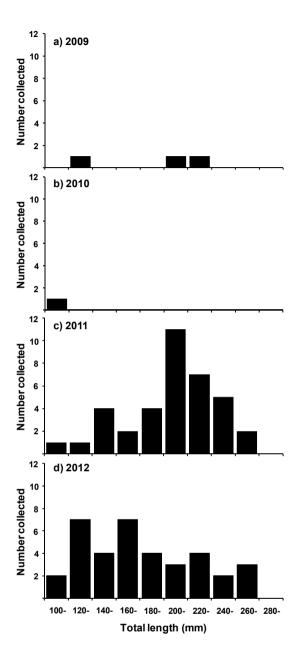


Figure 7. Length frequency of *Penaeus monodon* captured in the late spring and early summer (April – August) from 2009–2012.

For the full dataset, the total length\*sex interaction on WT was significant (P = 0.002), precluding us from examining TL (P < 0.001) and sex (P = 0.002) by themselves. For individuals between 150 and 250 mm TL, neither the effects of sex nor the total length\*sex interaction were significant (P = 0.081 and P = 0.072, respectively). This indicates that the relationship between WT and TL did not differ significantly

between males and females, although larger animals (TL > 250 mm) in our study tended to be female (Figure 9). Power curves resulted in a better fit than plotting quadratic equations to these data. Separate equations were determined for the sexes separately (Figure 9); however, given that statistical analysis did not find a difference in length-weight relationships between sexes for individuals between 150 and 250 mm, the equation for these pooled data was determined as:

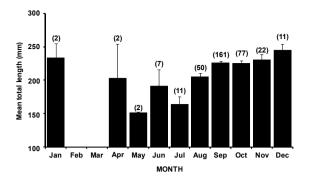
WT = 
$$7E^{-06} TL^{3.0401}$$
;  $r^2 = 0.918$ 

In our study, the largest female specimens were approximately 285 mm TL and 200 g, while the largest males were  $\sim$ 250 mm TL and  $\sim$ 140 g. Among the 191 *P. monodon* for which gender was definitively determined, the male to female ratio was 1.36:1.

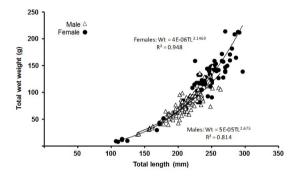
### Discussion

Reports of Penaeus monodon in the South Atlantic Bight and Gulf of Mexico have increased from 2008 levels. Some of the increase in the numbers reported in 2011 compared to earlier years may be attributable to greater efforts to document their occurrence; however, the magnitude of the increase suggests it is more than an artifact of this expanded reporting effort. Based on information from fisherman, the reduction in reports from 2011 to 2012, was probably due to "reporting apathy", as the novelty of reporting this species diminished. A similar phenomenon was observed for Indo-Pacific lionfish, Pterois volitans/miles (Linnaeus, 1758/Bennett, 1828) (Ruttenberg et al. 2012). Considering the increase, it is probable that a breeding population(s) is present, either in the South Atlantic Bight, the Gulf of Mexico, or both. In addition, the size range of individuals collected (102 mm to 298 mm TL) documents the presence of both juveniles and adults in the coastal waters of the U.S.

The geographic patterns in the number of *P. monodon* captured may be influenced by differences in commercial fishing effort among those areas and not entirely reflective of the abundance or distribution of the species. For example, *P. monodon* postlarvae typically recruit into estuaries from offshore breeding grounds in spring/summer, followed by their growth through early sub-adult stages in these shallow nursery grounds and subsequent offshore egress as observed within its native range (Motoh 1981).



**Figure 8.** Monthly mean length of *Penaeus monodon* collected from 2009–2012. Numbers shown in parentheses are sample sizes; error bars represent 1 S.E. of the mean.



**Figure 9.** Length-weight scatter plot of female and male *Penaeus monodon* captured from 2009–2012 (open triangles: males, n = 117; closed circle: females, n = 80).

This pattern of migration would provide inland access to *P. monodon* with a wider variety of fishing gear earlier in the year than can be successfully employed to capture adult *P. monodon* once they have moved offshore.

Introductions of *P. monodon* into the southeastern U.S. have three potential sources: 1) the release of larvae in ballast water taken onboard within their native range; 2) migration from areas in the Atlantic or Caribbean Sea where wild populations have become established (most likely as a result of prior aquaculture escape); and 3) escape from active and ongoing aquaculture facilities in the western Atlantic. Individuals found in U.S. waters may have arrived as a result of a continuous supply of *P. monodon* transported by currents from aquaculture facilities or wild populations in the Caribbean Sea or coastal South America.

Among the many vectors that can facilitate the introduction of nonindigenous marine and estuarine invertebrate species, ballast water release, along with hitchhikers in hull-fouling communities, are the most prominent (Carlton 2011). Chu et al. (1997) and G. Ruiz (Smithsonian Environmental Research Center, Edgewater, MD, pers. comm.) report having recovered viable larval decapod crustaceans from ballast water, and numerous crab species are believed to have been transferred between geographic regions via ballast water. This may have been the vector for the introduction from Asia to the eastern U.S. of the Oriental shrimp, Palaemon macrodactylus, and the Asian shore crab, Hemigrapsus sanguineus (De Haan, 1853) (G. Ruiz, pers. comm.). Although they recognized the uncertainty involved in determining the source of P. monodon introduced to Brazil, Severino-Rodrigues et al. (2000) hypothesized that ballast water discharge may have played a role in its introduction there.

Lessepsian transport of penaeid shrimps (i.e., from their native range into the Mediterranean via the Suez Canal; Rodriguez and Suárez 2001) and migration from areas where they have been introduced (e.g., along the South American coast-line; Pérez Farfante and Kensley 1997; Cintra et al. 2011; Leão et al. 2011) are also significant means of their dispersal. Altuve et al. (2008) suggested that the natural migration of *P. monodon*, assisted by the Guiana Current, spread the species from northern Brazil into the Caribbean, a distance of nearly 1600 km, in about four years.

Hurricanes and tropical storms frequently take a path through the Caribbean northward into the South Atlantic Bight or more westerly into the Gulf of Mexico. Tropical Storm Noel dropped 38-51 cm of rain along the southern coast of the Dominican Republic in 2007, where floodwaters came within 0.3 meters of pond berms at a facility where Litopenaeus vannamei was being cultured, and a massive escape was narrowly avoided (D. Drennan, Industria Nacional Agropesquera, Dominican Republic, pers. comm.). Those same ponds had previously been breached by flooding during periods when P. monodon was being cultured. In subsequent years, when L. vannamei was again being farmed, P. monodon larvae or postlarvae occasionally made their way into the ponds during post-harvest re-filling, indicating that a breeding population existed in nearby waters (D. Drennan, pers. comm.). Hurricanes Earl (2010) and Irene (2011) took similar tracks, providing more recent opportunities for storm-related transport to carry *P. monodon* into U.S. waters from the Caribbean. There is additional anecdotal information that suggests that shrimp farms in the Caribbean often lose portions of their crop during storm flooding, pond maintenance, or harvest operations (D. Knott, pers. comm.). Similar routine escape associated with shrimp pond harvest has been documented elsewhere (Wenner and Knott 1992).

An accidental release of *P. monodon* into the South Atlantic Bight, where they remained undetected for 18 years following their 1988 escape, is an unlikely explanation for their recent appearance in the U.S. Although commercial shrimpers caught them in the two months following their release (August-September 1988; D. Knott, pers. comm.), there were no subsequent reports until 2006. Considering the intense fishing efforts of the shrimp trawling industry along the entire southeastern U.S. coastline, along with the estuarine nursery habitat of *P. monodon*, it is improbable that a population became established there following the 1988 release and avoided detection until 2006.

Carlton (2011) disputed the notion that introduced species seldom have a notable impact in the newly colonized communities (e.g., see Vander Zanden 2005), declaring that for up to 95% of the known marine crustacean invasions there is a dearth of qualitative, quantitative, or experimental studies of ecological or other impacts. In light of this information gap, Ruiz et al. (2011) assembled a summary of historical records of marine crustacean invasions in North America and concluded that the negative ecological or economic impacts have only been reported for 28% of the 108 nonindigenous crustaceans found in marine, estuarine, and freshwater habitats of that continent. Furthermore, even for that 28%, the impacts were poorly documented. Not surprisingly then, the impacts of P. monodon on native fauna in areas where it has been introduced currently poorly documented. Because are *P. monodon* feeds on benthic organisms, primarily small crabs, shrimp, bivalves, and gastropods (Marte 1980), direct predation on local fauna is a concern. In addition to the potential effects of predation, the larger size of P. monodon may also confer greater nutritional requirements and a competitive advantage over native species in obtaining food.

The continual emergence, discovery, and global spread of novel shrimp pathogens (Flegel 2012; Lightner et al. 2012) pose the potential for unpredictable impacts on native crustacean

fauna. Although the mortality caused by many of these pathogens is generally greater in culture facilities than in the wild, the impact of infected seed and broodstock transfers on wild shrimp populations is poorly known. In 2003, the Pacific whiteleg shrimp, Litopenaeus vannamei (Boone, 1931) eclipsed P. monodon in terms of global culture production, largely due to the susceptibility of P. monodon to a variety of viral diseases (Spaargaren 1996; Flegel 1997; Primavera and Quinitio 2000; Flegel 2012), with white spot syndrome virus (WSSV) considered the most deleterious (Flegel and Alday-Sanz 1998; Global Aquaculture Alliance 1999; Soowannayan and Phanthura 2011). Penaeus monodon is capable of transmitting viral diseases such as WSSV to native shrimp species and other crustaceans (Chou et al. 1998; Kanchanaphum et al. 1998; Soowannayan and Phanthura 2011). The most recent threat to cultured P. monodon and L. vannamei in Asia, acute hepatopancreatic necrosis syndrome (AHPNS), was discovered in China in 2009 and has since caused widespread shrimp mortality on farms throughout southeast Asia, including Vietnam, Malaysia, and Thailand (Flegel 2012; Leaño and Mohan 2012). This disease is an emerging idiopathic disease whose potential for transmission to wild populations of penaeid shrimp is virtually unknown.

We are currently working to identify the source(s) of Asian tiger shrimp along the Gulf and Atlantic coasts of the U.S., with the hope of answering some of the questions regarding their establishment, the number of introductions, and their population structure. Preliminary phylogenetic analyses have indicated little genetic diversity in samples collected along the coast from North Carolina through the Gulf of Mexico. This lack of genetic differentiation indicates that founding individuals are highly related or inbred as a result of related stocks or genetic bottlenecks in the aquaculture industry. Further studies using additional markers and Hardy-Weinberg linkage equilibrium assessments could assist in determining whether P. monodon populations are established and reproducing in the region. With adequate samples and genetic markers, information can be gleaned on the populations of origin, number of founding populations, and dispersal pathways in the U.S. Samples examined to date are dissimilar to published mitochondrial genome and voucher specimen sequences of animals collected in their native range.

We propose that *P. monodon* is established in the southeastern U.S. along the Atlantic coast

and in the Gulf of Mexico. This is suggested by the overall increase in specimen collections, as well as the occurrence of individuals that span a range of sizes (juvenile to adult) and the increasing number of reports from inshore areas that are typically considered to be nursery grounds for many penaeid species. Ongoing genetic studies are being conducted to verify this establishment, including the development of a genetic probe to conduct environmental-DNA (eDNA) analyses for the detection of P. monodon at sizes that are not typically captured by trawl, seine, or cast nets. Such analyses could be applied to plankton samples to see whether or not they contain P. monodon larval stages. Specific nursery areas have yet to be identified, and these genetic approaches are likely to assist in this regard. Once these are located, a comparison of the temperature regime of the South Atlantic Bight with that required to support reproduction by this species is needed, as is a determination of the seasonality of reproduction in its newly introduced range. Neither the impacts on native fauna of this recently arrived shrimp species nor the source(s) or vector(s) of the invasion are known at this time. Further research on the sources of this introduction, the locations of breeding populations, and the ecological consequences of this invasion are warranted.

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### References

- Aguado NG, Sayegh J (2007) Presencia del camarón tigre gigante Penaeus monodon (Crustacea, Penaeidae) en la costas del Estado Anzoátegui, Venezuela. Boletim do Instituto Oceanográfico, Venezuela 46: 107–111
- Altuve DE, Marcano LA, Alió JJ, Blanco-Rambla JP (2008) Presencia del camarón tigre *Penaeus monodon* (Fabricius, 1798) en la costa del delta del río Orinoco y golfo de Paria, Venezuela. *Memoria de la Fundación La Salle de Ciencias Naturales* 68(169): 83–92
- Anyanwu PE, Ayinla OA, Ebonwu BI, Ayaobu-Cookey IK, Hamzat MB, Ihimekpen AF, Matanmi MA, Afolabi ES, Ajijo MR, Olaluwoye BL (2011) Culture possibilities of *Penaeus* monodon. Nigerian Fisheries and Aquatic Sciences 6: 499– 505
- Ayinla OA, Anyanwu PE, Solarin BB, Hamzat B, Ebonwu BI et al. (2009) Collection and maturation of broodstock of black tiger shrimp, *Penaeus monodon*, in Nigeria. Proceedings of the 24th Annual Conference of the Fisheries Society of Nigeria (FISON), October 25-28, 2009, Nigeria, pp 91–95
- Beard TW, Wickins JF (1980) Breeding of *Penaeus monodon* Fabricius in laboratory recirculation systems. *Aquaculture* 20(2): 79–89, http://dx.doi.org/10.1016/0044-8486(80)90037-X
- Carlton JT (2011) The global dispersal of marine and estuarine crustaceans. In: Galil BS, Clark PF, Carlton JT (eds), In the wrong place alien marine crustaceans: distribution, biology and impacts, invading nature. *Springer Series in Invasion Ecology* 6: 3–23
- Chaudhari KJ, Jalihal DR (1993) A field key to the seed penaeid prawns along the Konkan Coast (west coast of India). *Crustaceana* 65: 318–335, http://dx.doi.org/10.1163/156854093 X00757
- Chou HY, Huang CY, Lo CF, Kou GH (1998) Studies on transmission of white spot syndrome associated baculovirus (WSBV) in *Penaeus monodon* and *P. japonicus* via waterborne contact and oral ingestion. *Aquaculture* 164: 263–276, http://dx.doi.org/10.1016/S0044-8486(98)00192-6
- Chu KH, Tam PF, Fung CH, Chen QC (1997) A biological survey of ballast water in container ships entering Hong Kong. *Hydrobiologia* 352: 201–206, http://dx.doi.org/10.1023/A:1003 067105577
- Cintra IHA, Paiva KS, Botelho MN, Silva KCA (2011) Presence of *Penaeus monodon* in the continental shelf of the State of Para, northern Brazil (Crustacea, Decapoda, Penaeidae). *Revista de Ciencias Agrarias* 54(3): 314–317, http://dx.doi.org/ 10.4322/rca.2012.028
- Coelho PA, Santos MCF, Ramos-Porto M (2001) Ocorrência de Penaeus monodon Fabricius, 1798 no litoral dos estados de Pernambuco e Alagoas (Crustacea, Decapoda, Penaeidae). Boletim Técnico Cienífico CEPENE, Tamandaré 9: 149–153
- Crosnier A (1965) Les crevettes penaeides du plateau continental Malgache. *Cahiers ORSTOM (Océanographie)* 3, Suppl.3: 158 pp
- Dall W (1992) Feeding, digestion and assimilation in Penaeidae.
  In: Allan GL, Dall W (eds), *Proceedings of the Aquaculture Nutrition Workshop*, Salamander Bay, Australia, 15–17 April 1991. NSW Fisheries, Brackish Water Fish Culture Research Station, Salamander Bay, NSW, Australia, pp 57–63
- Delmendo MN, Rabanal HR (1956) Cultivation of "sugpo" (jumbo tiger shrimp), *Penaeus monodon* Fabricius, in the Philippines. *Proceedings of the Indo-Pacific Fisheries Council* 6(2–3): 424–431
- FAO (2012) Species Fact Sheets: *Penaeus monodon* (Fabricius, 1798). In: FAO Fisheries and Aquaculture Department. http://www.fao.org/fishery/species/3405/en (Accessed 29 May 2013)

- FAO (2013) Cultured Aquatic Species Information Programme: *Penaeus monodon* (Fabricius, 1798). In: FAO Fisheries and Aquaculture Department. http://www.fao.org/fishery/cultured species/Penaeus\_monodon/en (Accessed 29 May 2013)
- Flegel TW (1997) Special topic review: Major viral diseases of the black tiger prawn (*Penaeus monodon*) in Thailand. World Journal of Microbiology and Biotechnology 13: 433–442, http://dx.doi.org/10.1023/A:1018580301578
- Flegel TW, Alday-Sanz V (1998) The crisis in Asian shrimp aquaculture: current status and future needs. Journal of Applied Ichthyology – Zeitschrift Für Angewandte Ichthyologie 14: 269–273
- Flegel TW (2012) Historic emergence, impact and current status of shrimp pathogens in Asia. *Journal of Invertebrate Pathology* 110: 166–173, http://dx.doi.org/10.1016/j.jip.2012.03.004
- Global Aquaculture Alliance (1999) Shrimp white spot virus disease in Latin America an update. *Global Aquaculture Advocate* 2: 34–45
- Global Biodiversity Information Facility (2013) Biodiversity occurrence data published by: Senckenberg: Collection Crustacea - ZMB (Accessed through Global Biodiversity Information data portal, http://data.gbif.org/datasets/resource/11852, 29 May 2013)
- Gómez-Lemos LA, Campos NH (2008) Presencia de *Penaeus* monodon Fabricius (Crustacea: Decapoda: Penaeidae) en aguas de la Guajira colombiana. Boletim Investigacao Maritíma, Costeras 23: 221–225
- Hall DNF (1962) Observations on the taxonomy and biology of some Indo-West Pacific Penaeidae (Crustacea, Decapoda). Fishery Publications, Colonial Office, 17, pp 1–229
- Hill BJ, Wassenberg TJ (1987) Fate of discards from prawn trawlers in Torres Strait. *Australian Journal of Marine and Freshwater Research* 41: 51–64
- Jintoni B (2003) Water quality requirements for *Penaeus monodon* culture in Malaysia. Department of Fisheries Technical Reference No. 2, Department of Fisheries, Sabah, Malaysia. http://www.fishdept.sabah.gov.my/techrefs.asp (Accessed 29 May 2013)
- Kanchanaphum P, Wongteerasupaya C, Sitidilokratana N, Boonsaeng V, Panyim S, Tassanakajon A, Withyachumnarnkul B, Flegel TW (1998) Experimental transmission of white spot syndrome virus (WSSV) from crabs to shrimp *Penaeus monodon. Diseases of Aquatic Organisms* 34: 1–7, http://dx.doi.org/10.3354/dao034001
- Leaño EM, Mohan CV (2012) Early mortality syndrome threatens Asia's shrimp farms. *Global Aquaculture Advocate* July/August 2012: 38–39
- Leão TCC, Almeida WR, Dechoum M, Ziller SR (2011) Espécies exóticas invasoras no Nordeste do Brasil: contextualização, manejo e políticas públicas. CEPAN, Instituto Hórus, Recife, 99 pp
- Lightner DV, Redman RM, Pantoja CR, Tang KFJ, Noble BL, Schofield P, Mohney LL, Nunan LM, Navarro SA (2012) Historic emergence, impact and current status of shrimp pathogens in the Americas. *Journal of Invertebrate Pathology* 110: 174–183, http://dx.doi.org/10.1016/j.jip.2012.03.006
- Lumare F, Di Muro P, Tenderini L, Zupo V (1993) Experimental intensive culture of *Penaeus monodon* in the cold-temperate climate of the north-east coast of Italy (a fishery 'valle' of the River Po Delta). *Aquaculture* 113: 231–241, http://dx.doi.org/ 10.1016/0044-8486(93)90476-F
- Luna-Marte C (1982) Seasonal variation in food and feeding of Penaeus monodon Fabricius (Decapoda, Natantia). Crustaceana 42: 250–255, http://dx.doi.org/10.1163/156854082 X00957
- Marte CL (1980) The food and feeding habit of *Penaeus monodon* Fabricius collected from Makato River, Aklan, Philippines (Decapoda Natantia). *Crustaceana* 38: 225–236, http://dx.doi.org/10.1163/156854080X00139

- Menasveta P, Sangpradub S, Piyatiratitivorakul S, Fast AW (1994) Effects of broodstock size and source on ovarian maturation and spawning of *Penaeus monodon* Fabricius from the Gulf of Thailand. *Journal of the World Aquaculture Society* 25(1): 41–49, http://dx.doi.org/10.1111/j.1749-7345.1994. tb00802.x
- Mohamed KH (1967) Synopsis of biological data on the jumbo tiger prawn *Penaeus monodon* Fabricius 1798. Species Synopsis No. 3. FAO World Scientific Conference on the Biology and Culture of Shrimps and Prawns. Ciudad de Mexico 12-24/6/1967. http://www.fao.org/docrep/005/AC765T/ AC765T00.htm (Accessed 30 January 2014)
- Motoh H (1981) Studies on the fisheries biology of the giant tiger prawn, *Penaeus monodon* in the Philippines. Technical Report No. 7, SEAFDEC, Philippines, 128 pp
- Motoh H (1985) Biology and ecology of Penaeus monodon. In: Taki Y, Primavera JH, Llobrera JA (eds), Proceedings of the First International Conference on the Culture of Penaeid Prawns/Shrimps, 4-7 December 1984, Iliolo City, Philippines. Aquaculture Department, Southeast Asian Fisheries Development Center, pp 27–36
- Motoh H, Buri P (1980) Development of the external genitalia of the giant prawn, *Penaeus monodon. Bulletin of the Japanese* Society of Scientific Fisheries 46(2): 149–155, http://dx.doi.org/ 10.2331/suisan.46.149
- NOAA (National Oceanic and Atmospheric Administration) (2013) National Marine Fisheries Service, Monthly Commercial Landings Statistics http://www.st.nmfs.noaa.gov/stl/ commercial/landings/monthly\_landings.html (Accessed 2 June 2013)
- Pérez Farfante I, Kensley B (1997) Penaeoid and sergestoid shrimps and prawns of the world – keys and diagnosis for the families and genera. Mémoires du Museum National d'historoire Naturelle 5: 1–233
- Primavera JH (1982) Studies on broodstock of sugpo *Penaeus* monodon Fabricius and other penaeids at the SEAFDEC Aquaculture Department. Marine Biological Association of India, Proceedings of the Symposium on Coastal Aquaculture, Cochin, India, 12-18 January 1980, 1, pp 28–36
- Primavera JH, Parado-Estepa FD, Lebata JL (1998) Morphometric relationship of length and weight of giant tiger prawn *Penaeus monodon* according to life stage, sex and source. *Aquaculture* 164: 67–75, http://dx.doi.org/10.1016/S0044-8486 (98)00177-X
- Primavera JH, Quinitio ET (2000) Runt-deformity syndrome in cultured giant tiger prawn *Penaeus monodon*. Journal of Crustacean Biology 20(4): 796–802
- Rodríguez G, Suárez H (2001) Anthropogenic dispersal of decapod crustaceans in aquatic environments. *Interciencia* 26(7): 282–288
- Ruiz G, Fofonoff P, Steves B, Dahlstrom A (2011) Marine crustacean invasions in North America: a synthesis of historical records and documented impacts. In: Galil BS, Clark PF, Carlton JT (eds), In the wrong place – alien marine crustaceans: distribution, biology and impacts, invading nature. Springer Series in Invasion Ecology 6, pp 215–250
- Ruttenberg BI, Schofield PJ, Akings JL, Acosta A, Feeley MW, Blondeau J, Smith SG, Ault JS (2012) Rapid invasion of Indo-Pacific Lionfishes (*Pterois volitans* and *Pterois miles*) in the Florida Keys, USA: evidence from multiple pre- and post-invasion data sets. *Bulletin of Marine Science* 88(4): 1051–1059, http://dx.doi.org/10.5343/bms.2011.1108
- Sahel and West Africa Club (2006) Exploring Economic Opportunities in Sustainable Shrimp Farming in West Africa: Focus on South-South Cooperation. Meeting Report. Organization for Economic Co-operation and Development. http://www.oecd.org/dataoecd/56/2/38524451.pdf (Accessed 29 May 2013)

- Severino-Rodrigues E, Barreto OJS, Perroni RW (2000) Penaeus monodon Fabricius (Crustacea, Decapoda, Penaeidae) no estuário de Santos. In: I Congresso Brasileiro sobre Crustaceos, 16-20 October 2000. Resumos, São Pedro, Brazil, pp 186
- Silva KCA, Ramos-Porto M, Cintra IHA (2002) Registro de Penaeus monodon Fabricius, 1798, na plataforma continental do estado do Amapá (Crustacea, Decapoda, Penaeidae). Boletim Técnico Cientifico Centro de Pesquisa e Gestão de Recursos Pesqueiros do Litoral Norte (CEPNOR), Belém 2(1): 75–80
- Smith DM, Dall W, Moore LE (1992) The natural food of some Australian penaeids. In: Allan GL, Dall W (eds), Proceedings of the Aquaculture Nutrition Workshop, Salamander Bay, Australia 15–17 April 1991. NSW Fisheries, Brackish Water Fish Culture Research Station, Salamander Bay, NSW, Australia, pp 95–96
- Soowannayan C, Phanthura M (2011) Horizontal transmission of white spot syndrome virus (WSSV) between red claw crayfish (*Cherax quadricarinatus*) and the giant tiger shrimp *Penaeus monodon. Aquaculture* 319: 5–10, http://dx.doi.org/10. 1016/j.aquaculture.2011.06.012
- Spaargaren DH (1996) Diseases in cultures of tiger prawns, *Penaeus monodon* Fabricius (Decapoda, Natantia). *Crustaceana* 69: 1018–1024, http://dx.doi.org/10.1163/1568540 96X00457

- Srivatsa KR (1953) A survey and comparative analysis of prawn (shrimp) fishery of the Gulf of Kutch in Saurashtra in Western India. Government of Saurashtra, India, Department of Industries and Supplies
- Thomas MM (1972) Food and feeding habits of *Penaeus* monodon Fabricius from Korapuzha estuary. *Indian Journal* of Fisheries 19: 202–204
- USGS (U.S. Geological Survey) (2013) Nonindigenous aquatic species database. Gainesville, FL. http://nas.er.usgs.gov/que ries/factsheet.aspx?speciesID=12097 (Accessed 29 May 2013)
- Vander Zanden MJ (2005) The success of animal invaders. Proceedings of the National Academy of Sciences 102(20): 7055–7056, http://dx.doi.org/10.1073/pnas.0502549102
- Wenner EL, Knott DM (1992) Occurrence of Pacific white shrimp, *Penaeus vannamei*, in coastal waters of South Carolina. In: DeVoe HR (ed), Introductions and transfers of marine species: achieving a balance between economic development and resource protection. Proceedings of the Conference and Workshop, 30 October – 2 November 1991, Hilton Head Island, South Carolina. South Carolina Sea Grant Consortium, Charleston, SC, pp 173–191